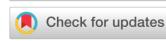
# Natural Resources for Human Health



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# Potential of Streptomyces in producing antiplasmodial lead compounds

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ABSTRACT: Streptomyces are bacteria of great importance for several decades. Numerous potent metabolites characterized as antibiotics including macrolides and polypetides have been reported from Streptomyces and developed as effective drugs for the treatment of several illnesses. Therefore, Streptomyces can be considered as an important source of bioactive compounds which might help in the eradication of malaria which remains one of the greatest threats to human life, especially in the tropical and sub-tropical regions. The reported in vitro antiplasmodial properties of chemical constituents from Streptomyces strains have led to promising results like bafilomycin A1 (9), concanamycin A (10), elaiophylin (17), cyclomarin C (23), urdamycinone E (44), geldanamycin (52) and metacycloprodigiosin (74) which individually exhibited strong antiplasmodial activity against the chloroquine-resistant strain *Plasmodium falciparum* K1 with IC<sub>50</sub> values of 0.041  $\mu$ g/ml, 0.2 nM, 0.22  $\mu$ g/ml, 0.24  $\mu$ g/ml, 0.0534  $\mu$ g/ml, 0.35  $\mu$ g/ml and 0.0050  $\mu$ g/ml, respectively. In some cases, the tested compound was most active than the reference and without observed toxicity until the highest concentration. However, more in vivo and toxicity studies are necessary for further guidance in the process of drug development. To the best of our knowledge, no specific review has been done on the potential of Streptomyces in furnishing antiplasmodial compounds for malaria control. This paper aims to compile the literature up to 2021 on antiplasmodial compounds isolated from Streptomyces for easy and rapid access to the literature for further investigations in continuity.

# 1. INTRODUCTION

The last report of the World Health Organisation (WHO) on malaria worldwide indicates that there was an increased case incidence of malaria between 2020 and 2021 due to the disruption to services during the Covid-19 pandemic with almost 241 million malaria cases and 627,000 deaths in 85 malaria-endemic countries (WHO, 2021). observation supports that malaria is still among the most harmful parasitic diseases and a major public health problem in several tropic and subtropic regions of the world, especially the developing countries which are poorly deserved with equipped medical centres and where the resistance of Plasmodium falciparum to the prescribed antimalarial drugs have been reported (Nasomjai et al., 2014; Ogbole et al., 2018). Plasmodium falciparum is the most virulent parasite that causes the most severe forms of malaria and the highest rate of mortality with children and pregnant women as the most affected people (Júnior et al., 2012). Several strategies have been proposed by WHO in preventing and curing the diseases over the last decades, including the combination of effective therapeutic agents like artemisinin-based combination therapy (ACT), the large distribution of long-lasting insecticidetreated bed nets or the development of new insecticides to eliminate its malaria mosquito vectors (Happi et al., 2015, 2022). However, new more potent antimalarial drugs are needed to supply the ones existing already and address the observed resistance (Bathurst & Hentschel, 2006). 2006, four main classes of compounds have been identified among the most prescribed antimalarial drugs: The quinine derivatives like chloroquine, mefloquine, amodiaquine or the aminoquinolines like primaquine; the antifolate compounds like pyrimethamine, dapsone or sulfadoxine, the artemisinin derivatives like artesunate, artemether or co-Artem; and finally the hydroxynaphthoquinone like atovaquone (Baniecki et al., 2007). In this regard for new antiplasmodial drug discovery, it seems imperative to explore new sources of bioactive metabolites

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with new structural diversity to increase the opportunities in structure modifications as well as in the development of a high number of possibilities of active compounds.

The bacteria called Streptomyces represent an interesting source of specialized metabolites with a high structural diversity and a large scale of biological activities. It is well reported that mostly produced antibiotics like chloramphenicol from Streptomyces venezuelae and used in the treatment of typhoid, rifampicin and vancomycin from S. mediterranei and S. orientalis, respectively, and which have been important antibiotics prescribed in the treatment of leprosy and methicillinresistant Staphylococcus aureus, respectively (Chater, 2006). Many of these antibiotics derived from Streptomyces are highly functionalised compounds that mostly belong to the class of macrolides or cyclopolypeptides. Like other microorganisms (fungi or bacteria) which live in their hosts (Happi et al., 2015), Streptomyces can be isolated from plant material, soil or marine sources like sea plant, sea sediment or sea animals. Our literature survey provided us with significant data on the high structural diversity and strong activity of some Streptomyces compounds compared to the well-known standard drugs and which deserve further attention in the development of new potent antimalarial drugs. To the best of our knowledge, no review article has been published on the phytochemistry and pharmacology of the bacteria Streptomyces for their contribution in fighting against malaria as a source of antiplasmodial agents. This review covers the documented works up to December 2021.

# 2. METHODOLOGY

This paper has been written based on collected data previously reported in the literature over the last decades up to 2021. Numerous online libraries including Scifinder and Scifinder<sup>n</sup>, PubMed, Google Scholar as well as Web of Science were used in searching for information on antiplasmodial metabolites from *Streptomyces*. The keywords *Streptomyces*, *Plasmodium*, antiplasmodial and malaria were used to monitor and refine our search without language restriction.

# 3. ANTIPLASMODIAL LEAD COMPOUNDS FROM STREPTOMYCES

The literature survey on the previous chemical and pharmacological studies of *Streptomyces* revealed that numerous specialized metabolites, which are mostly antibiotics, have been isolated from an important number of strains that were not fully identified in many cases but reported as *Streptomyces* sp. and their original strain codes from the authors' databank. The reported *Streptomyces* sp. have been isolated from plant material like rice (Supong, Thawai, et al., 2016), from marine source (Buedenbender et al., 2018; Supong et al., 2012) or from soil (Intaraudom et al., 2015; Supong, Sripreechasak, et al., 2016). Among the reported compounds from *Streptomyces*, 70 distinct compounds have demonstrated an antiplasmodial potency including 51 natural products (~72.9 %) and 19 synthetic derivatives (~27.1 %) prepared from isolated natural products. The 51 natural occurring antiplasmodial compounds

from *Streptomyces* can be organized into thirteen different classes of specialized metabolites sorted as follows: Macrolides were the most abundant (~23.5 %) followed by anthraquinones (~15.7 %), polypeptides (~11.7 %), geldanamycin analogues (~7.8 %), pactamycin analogues, polyether compounds and diketopiperazines (~5.9 %, each), cyclopeptides and carbazomycin analogues (~3.9 %, each), sesquiterpenoid, diterpenoid and zeatin-type compound (~2.0 %, each) and others (~9.8 %). As we recently reported, a pure compound should exhibit an  $IC_{50} \le 10 \ \mu M$  to be considered as an active compound by the industry (Happi et al., 2022).

Furthermore, the seventy reported antiplasmodial compounds from Streptomyces have been tested against a total of eight Plasmodium falciparum strains including four chloroquine-sensitive strains (P. falciparum 3D7, D6, NF54 and HB3) and four chloroquine-resistant strains (P. falciparum K1, Dd2, W2 and 7G8) (Table 1). Globally, the resistant strains K1, Dd2 and the sensitive strain 3D7 have been the most used during the antiplasmodial tests of Streptomyces compounds. Thus, the promising results against the resistant strains indicated that Streptomyces can be an important source of active principles to handle the resistance of P. falciparum which represent the main issue in controlling the progress and the eradication of malaria (Happi et al., 2022). Artemisinin (1) and its derivatives dihydroartemisinin (2) and artesunate (3), as well as the gold antimalarial drug chloroquine (4) and other standard drugs like mefloquine (5), puromycin (6), pyrimethamine (7) and pyronaridine (8), have been used as reference compounds (standards) during the reported investigations (Figure 1). Their potencies against the used strains of P. falciparum are compiled in Table 1.

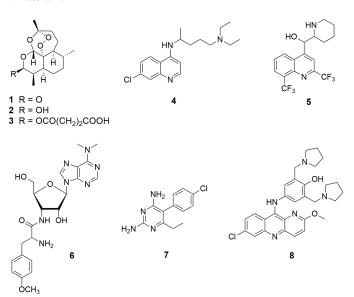


Figure 1. Some drugs used as standard for antiplasmodial tests.

The reported active compounds from *Streptomyces* can be classified into five categories based on their potencies in comparison with the criteria for antiplasmodial activity against *P. falciparum* used by the World Health Organization that



 Table 1

 Potencies of standard drugs used during the recorded antiplasmodial tests.

| Type of P. falciparum | Strains index | Standard, IC 50                 | Reference                     |
|-----------------------|---------------|---------------------------------|-------------------------------|
|                       | 3D7           | Artesunate (3), 0.9 nM          | Kiefer et al. (2019)          |
|                       |               | Chloroquine (4), 3.4 nM         | Buedenbender et al. (2018)    |
|                       |               | Dihydroartemisinin (2), 0.4 nM  |                               |
|                       |               | Puromycin (6), 148.9 nM         |                               |
| CQ-sensitive strains  |               | Pyrimethamine (7), 4.7 nM       |                               |
|                       |               | Pyronaridine (8), 7.4 nM        |                               |
|                       | D6            | Chloroquine (4), 10.6 nM        | Almabruk et al. (2013)        |
|                       | NF54          | Chloroquine (4), 6 nM           | Happi et al. (2015)           |
|                       | HB3           | Chloroquine (4), 9.47 nM        | Baniecki et al. (2007)        |
|                       |               | Mefloquine (5), 9.63 nM         |                               |
|                       |               | Artemisinin (1), 9.70 nM        |                               |
|                       | K1            | Artemisinin (1), 3.9 nM         | Isaka et al. (2002)           |
|                       |               | Chloroquine (4), 0.46 uM        | Jang et al. (2017)            |
|                       |               | Dihydroartemisinin (2), 1.98 nM | Intaraudom et al. (2015)      |
|                       |               | Mefloquine (5), 29.1 nM         | Supong, Thawai, et al. (2016) |
|                       | Dd2           | Artesunate (3), 1.3 nM          | Buedenbender et al. (2018)    |
| CQ-resistant strains  |               | Chloroquine (4), 87.9 nM        |                               |
|                       |               | Dihydroartemisinin (2), 0.6 nM  |                               |
|                       |               | Puromycin (6), 114.4 nM         |                               |
|                       |               | Pyronaridine (8), 8.3 nM        |                               |
|                       | 7G8           | Chloroquine (4), 89.5 nM        | Almabruk et al. (2013)        |
|                       | W2            | Chloroquine (4), 70 nM          | Mackinnon et al. (1997)       |

CQ: chloroquine

indicates that : Pronounced activity (IC $_{50}$  < 5  $\mu g/ml$  or IC $_{50} \leq 0.1~\mu M)$ ; good activity (5 < IC $_{50}$  <10  $\mu g/ml$  or IC $_{50}$  > 0.1  $\mu M$  but  $\leq 5~\mu M)$ ; moderate activity (10 < IC $_{50}$  < 20  $\mu g/ml$  or IC $_{50}$  > 5  $\mu M$  but < 20  $\mu M)$ ; low activity (20 < IC $_{50}$  < 40  $\mu g/ml$  or IC $_{50}$  > 20  $\mu M$  but < 50  $\mu M)$  and inactive if IC $_{50}$  > 40  $\mu g/ml$  or IC $_{50}$  > 50  $\mu M$  (WHO 2022; ) (Happi et al., 2022).

# 3.1. Macrolides

Macrolides represent an important class of antibiotics isolated from microorganisms including the most famous ones azithromycin and erythromycin which are pharmaceutical drugs available on market. Through the last decades, the chemical investigations of several *Streptomyces* strains led to the isolation and characterization of a number of macrolides including nineteen which were identified as potent antiplasmodial agents according to the data available in the literature so far (Table 2, Figure 2).

Earlier in 2002, the bacterial strain *Streptomyces spectabilis* BCC4785 has been isolated from the soil sample and the bio-guided fractionation of its extract led to the isolation of the macrolide bafilomycin A1 (9) showing a significant activity (IC $_{50}$  = 0.041  $\mu$ g/ml) against *Plasmodium falciparum* K1 (Isaka et al., 2002). Later in 2006, another macrolide named concanamycin A (10) from *Streptomyces* sp. displayed a strong antiplasmodial activity supported by its IC $_{50}$  of 0.2 nM against *P. falciparum* K1 (Auparakkitanon & Wilairat, 2006). Two 20-membered macrolides samroiyotmycins A (11) and B (12) were reported from *Streptomyces* sp. BCC33756 and tested for their antiplasmodial potency which gave the IC $_{50}$  values of

3.65  $\mu$ g/ml and 3.16  $\mu$ g/ml, respectively, against *P. falciparum* K1 (Dramae et al., 2013).

In 2016, Supong and co-workers have isolated the strain BCC72023 of Streptomyces sp. from Oryza sativa (rice). Its screening for antiplasmodial compounds gave the identification of three macrolides namely efomycin M (13) reported for the first time from a natural source, efomycin G (14) and oxohygrolidin (15) which displayed interesting potency against P. falciparum K1 with IC<sub>50</sub> values of 5.23  $\mu$ g/ml, 2.37  $\mu$ g/ml and 2.30  $\mu$ g/ml, respectively (Supong, Thawai, et al., 2016). Within the same year, the same authors reported five additional macrolides from the terrestrial *Streptomyces* sp. BCC71188 including monoglycosylelaiolide (16), elaiophylin also called azalomycin (17), 11,11'-odimethylelaiophylin (18), oxohygrolidin (19) and the previously-reported efomycin G (14) which displayed significant antiplasmodial activity against *P. falciparum* K1 with IC<sub>50</sub> values of 2.46  $\mu$ g/ml, 0.22  $\mu$ g/ml,  $1.47 \mu g/ml$ ,  $2.30 \mu g/ml$  and  $2.37 \mu g/ml$ , respectively (Supong, Sripreechasak, et al., 2016).

Most recently in 2018, elaiophylin (**20**) has been reported from *Streptomyces* sp. USC-16018 with an IC<sub>50</sub> of 777.9 nM with 96.6% inhibition at 40  $\mu$ M against the chloroquinesensitive strain *P. falciparum* 3D7 while it showed a more relevant activity against *P. falciparum* Dd2 with an IC<sub>50</sub> of 598.5 nM with 86.1% inhibition at 40  $\mu$ M (Buedenbender et al., 2018).

Furthermore, six macrolides have been partially characterized as high functionalized compounds which showed interesting antiplasmodial activity. Briefly, munumbicins A–D isolated



Figure 2. Antiplasmodial macrolides from Streptomyces.

**18**  $R_1 = R_2 = Me$ ,  $R_3 = A$ 

from *Streptomyces* NRRL30562 demonstrated strong antiplasmodial activity against *P. falciparum* CSC-1 (Honduras) with very low IC<sub>50</sub> values of 175 ng/ml, 130 ng/ml, 6.5 ng/ml and 4.5 ng/ml, respectively. More interestingly, the potency of munumbicin D was determined as almost 50% above that of the standard chloroquine and it did not show any observable lysis of human red blood cells up to a concentration of 80  $\mu$ g/ml which qualified munumbicin D as a good candidate for the development of new malarial drugs (Castillo et al., 2002). Moreover, the same authors characterized two other munumbicins (E-4 and E-5) from *Streptomyces* NRRL3052

A ŌH

which were less potent than the previous ones with IC<sub>50</sub> values of 0.50  $\mu$ g/ml and 0.87  $\mu$ g/ml, respectively, against the same strain of *P. falciparum* CSC-1 (Honduras) (Castillo et al., 2006).

# 3.2. Polypeptides

Besides the macrolides, another important class of antibiotics from natural sources is represented by the cyclopolypeptides also designated as polyketides. They consist of an association of several amino acids (or peptide) units which can be cyclic or acyclic. Depending on the number of units, we can distinguish



 Table 2

 Antiplasmodial macrolides from Streptomyces.

| N° | Name                        | Strain, IC 50        | Source                     | Reference   |
|----|-----------------------------|----------------------|----------------------------|---|
| 9  | Bafilomycin A1              | K1, 0.041 $\mu$ g/ml | S. spectabilis BCC 4785    | Isaka et al. (2002)   |
| 10 | Concanamycin A              | K1, 0.2 nM           | Streptomyces sp.           | Auparakkitanon and Wilairat (2006)                                  |
| 11 | Samroiyotmycin A            | K1, 3.65 $\mu$ g/ml  | Streptomyces sp. BCC33756  | Dramae et al. (2013)  |
| 12 | Samroiyotmycin B            | K1, 3.16 $\mu$ g/ml  | streptomyces sp. BCC55/30  | Diamae et al. (2013)  |
| 13 | Efomycin M                  | K1, 5.23 $\mu$ g/ml  | Streptomyces sp. BCC72023  | Supong, Thawai, et al. (2016)                                       |
| 14 | Efomycin G                  | K1, 2.37 $\mu$ g/ml  | Streptomyces sp. BCC72023  | Supong, Sripreechasak, et al. (2016); Supong, Thawai, et al. (2016) |
|    |                             |                      | Streptomyces sp. BCC71188  |   |
| 15 | oxohygrolidin               | K1, 2.30 $\mu$ g/ml  | Streptomyces sp. BCC72023  | Supong, Thawai, et al. (2016)                                       |
| 16 | Monoglycosylelaiolide       | K1, 2.46 $\mu$ g/ml  |                            |   |
| 17 | Elaiophylin (or azalomycin) | K1, 0.22 $\mu$ g/ml  | Streptomyces sp. BCC71188  | Supong, Sripreechasak, et al. (2016)                                |
| 18 | 11,11'-Odimethylelaiophylin | K1, 1.47 $\mu$ g/ml  | Streptomytes sp. BCC/1188  | Supong, Suprecentasak, et al. (2010)                                |
| 19 | Oxohygrolidin               | K1, 2.30 $\mu$ g/ml  |                            |   |
| 20 | Elaiophylin                 | 3D7, 777.9 nM        | Streptomyces sp. USC-16018 | Buedenbender et al. (2018)  |
|    |                             | Dd2, 598.5 nM        |                            |   |

different subclasses of polypeptides. For instance, the literature survey indicated that six natural occurring antiplasmodial polypeptides including two hexadepsipeptides (21 and 22), two heptadepsipeptides (23 and 24) and two octadepsipeptides (42 and 43) have been reported from  $\it Streptomyces$  strains (Table 3 , Figure 3). Furthermore, seventeen new synthetic analogues (25–41) with significant potencies have been prepared from desoxycyclomarin  $\it C$  (24).

Among the antiplasmodial hexadepsipeptides reported so far from Streptomyces, mollemycin A (21) is the first glycohexadepsipeptide-polyketide isolated from *Streptomyces* sp. CMBM0244 and demonstrating an exceptional potency against the drug-sensitive *P. falciparum* 3D7 and the multidrug-resistant P. falciparum Dd2 with IC50 values of 7 nM and 9 nM, respectively (Raju et al., 2014), whereas the recent works of Watson et al. (2021) supported that valinomycin (22) obtained from Streptomyces sp. PR3 displayed potent activity (IC<sub>50</sub> of 3.75 ng/ml) in a single test against *P. falciparum* NF54, while more interestingly, they found that the activity was increasing in a mixed test when valinomycin was mixed in different ratios with cyclic polypropylene glycols (cPPG). A series of tests revealed that when the cPPG fraction showed an activity of 1792 ng/ml, but the mixtures 22+cPPG in the fixed ratio 4:1, 3:2, 2:3 and 1:4 will display an increased potency of 1.86 ng/ml, 0.90 ng/ml, 0.75 ng/ml and 0.53 ng/ml, respectively. Their study supported that cPPG can significantly and synergistically improve in vitro the antiplasmodial potency of valinomycin (22) (Watson et al., 2021).

Cyclomarin C (23) and desoxycyclomarin C (24) are two natural heptadepsipeptides obtained from *Streptomyces* sp. BCC26924 and *Streptomyces* sp. CNB-982, respectively. Both compounds showed strong antiplasmodial activity indicated by the IC $_{50}$  value of 0.24  $\mu$ g/ml for cyclomarin C (23) against *P. falciparum* K1 and 39.8 nM for desoxycyclomarin C (24) against *P. falciparum* 3D7 (Intaraudom et al., 2011; Kiefer et al., 2019). Seventeen new analogues (25–41) of desoxycyclomarin C have been synthesised and tested against the strains 3D7 and

Dd2 of *P. falciparum*. In this regard, the potencies (IC<sub>50</sub> values) were ranging from 4.4 nM to 452.0 nM against *P. falciparum* 3D7 and from 6.5 nM to 421.5 nM against *P. falciparum* Dd2 (Table 3 ). The molecular docking performed by the authors suggested that the presence of the N'-methyltryptophan unit and the  $\gamma$ , $\delta$ -unsaturated side chain might play an important role in the improvement of the antiplasmodial potencies of the synthesised analogues (Kiefer et al., 2019).

Finally, two octadepsipeptides viz octaminomycins A (42) and B (43) were previously obtained from Streptomyces sp. RK85-270 and tested for their antiplasmodial activity against three P. falciparum strains including the drug-sensitive strain 3D7 and the two multidrug-resistant strains Dd2 and K1. The results showed that both compounds possessed the same activity against P. falciparum 3D7 with IC<sub>50</sub> of 1.5  $\mu$ M, while compound 42 was more active than 43 against the two other strains with IC<sub>50</sub> values of 1.6  $\mu$ M and 1.3  $\mu$ M for compound 42 against *P. falciparum* Dd2 and K1, respectively; while compound 43 displayed IC<sub>50</sub> values of 1.1  $\mu$ M and 0.85  $\mu$ M, respectively. Additionally, the two compounds were more pharmacologically interesting due to their no significant cytotoxicity against human cervical cancer cells (HeLa), human promyelocytic leukaemia cells (HL-60), mouse temperature-sensitive cdc2 mutant cells (tsFT210), and rat kidney cells that were infected with ts25 (srcts-NRK) at the highest concentration of 30  $\mu$ M (Jang et al., 2017).

# 3.3. Anthraquinones

Four C-glycosylated benz[ $\alpha$ ]anthraquinones urdamycinones E (44) and G (45), dehydroxyaquayamycin (46) as well as urdamycin E (47) were isolated from the marine *Streptomyces* sp. BCC45596 and displayed potent antiplasmodial activity against P. falciparum K1 with IC<sub>50</sub> values of 0.0534  $\mu$ g/ml, 0.142  $\mu$ g/ml, 2.93  $\mu$ g/ml and 0.173  $\mu$ g/ml, respectively (Supong et al., 2012). The comparison of the potency with their structural changes (Table 4 , Figure 4) suggested that the free angular hydroxy groups in their structures might play an important



Figure 3. Antiplasmodial polypeptides from Streptomyces.



39 R<sub>1</sub> = Prenyl, R<sub>2</sub> = i-propyl
 40 R<sub>1</sub> = t-pentyl, R<sub>2</sub> = i-propyl
 41 R<sub>1</sub> = H, R<sub>2</sub> = i-propyl

 Table 3

 Antiplasmodial polypeptides from Streptomyces.

| Mollemycin A   3D7, 7 nM   Dd2, 9 nM   Dd2, 9 nM   Sreptomyces sp. (CMBM0244)   Raju et al. (2014)  |             |                    |                     |                             |                          |
|---|-------------|--------------------|---------------------|-----------------------------|--------------------------|
| Dd2, 9 nM   NF54, 3.75 ng/ml   Streptomyces sp. PR3   Watson et al. (2021)  | $N^{\circ}$ | Name               | Strain, IC $_{50}$  |                             | Reference                |
| 22  | 21          | Mollemycin A       | 3D7, 7 nM           | Streptomyces sp. (CMBM0244) | Raju et al. (2014)       |
| 23         Cyclomarin C         K1, 0.24 μg/ml         Streptomyces sp. BCC26924         Intaraudom et al. (2011)           24         Desoxycyclomarin C         3D7, 39.8 mM         Streptomyces sp. CNB-982           25         Compound 25a         3D7, 9.0 nM         Synthetic analogue           26         Compound 25b         3D7, 4.4 nM         Synthetic analogue           27         Compound 25c         3D7, 4.8 nM         Synthetic analogue           28         Compound 25d         3D7, 13.4 nM         Synthetic analogue           29         Compound 25e         3D7, 28.1 nM         Synthetic analogue           30         Compound 28a         3D7, 34.4 nM         Synthetic analogue           31         Compound 28a         3D7, 57.6 nM         Synthetic analogue           32         Compound 28c         3D7, 355.7 nM         Synthetic analogue           33         Compound 28c         3D7, 355.7 nM         Synthetic analogue           34         Compound 28e         3D7, 314.2 nM         Synthetic analogue           35         Compound 34a         3D7, 47.9 nM         Synthetic analogue           36         Compound 34a         3D7, 47.9 nM         Synthetic analogue           37         Compound 34c         3D7, 452.0 nM   |             |                    | Dd2, 9 nM           |                             |                          |
| Desoxycyclomarin C   3D7, 39.8 nM   Streptomyces sp. CNB-982  | 22          | Valinomycin        | NF54, 3.75 ng/ml    | Streptomyces sp. PR3        | Watson et al. (2021)     |
| 25  | 23          | Cyclomarin C       | K1, 0.24 $\mu$ g/ml | Streptomyces sp. BCC26924   | Intaraudom et al. (2011) |
| Dd2, 12.9 nM   Synthetic analogue   | 24          | Desoxycyclomarin C | 3D7, 39.8 nM        | Streptomyces sp. CNB-982    |                          |
| 26         Compound 25b         3D7, 4.4 nM         Synthetic analogue           27         Compound 25c         3D7, 47.8 nM         Synthetic analogue           28         Compound 25d         3D7, 13.4 nM         Synthetic analogue           29         Compound 25e         3D7, 28.1 nM         Synthetic analogue           30         Compound 28a         3D7, 34.4 nM         Synthetic analogue           30         Compound 28b         3D7, 57.6 nM         Synthetic analogue           31         Compound 28c         3D7, 355.7 nM         Synthetic analogue           32         Compound 28c         3D7, 355.7 nM         Synthetic analogue           33         Compound 28d         3D7, 230.3 nM         Synthetic analogue           34         Compound 28e         3D7, 314.2 nM         Synthetic analogue           35         Compound 34a         3D7, 47.9 nM         Synthetic analogue           36         Compound 34a         3D7, 452.0 nM         Synthetic analogue           37         Compound 34a         3D7, 452.0 nM         Synthetic analogue           37         Compound 35a         3D7, 303.5 nM         Synthetic analogue           38         Compound 35a         3D7, 303.5 nM         Synthetic analogue  | 25          | Compound 25a       | 3D7, 9.0 nM         | Synthetic analogue          |                          |
| Dd2, 6.5 nM   Synthetic analogue   Dd2, 76.0 nM   Dd2, 76.0 nM   Synthetic analogue   Dd2, 76.0 nM   Dd2, 17.8 nM   Dd2, 17.8 nM   Synthetic analogue   Dd2, 17.8 nM   Dd2, 17.8 nM   Synthetic analogue   Dd2, 27.5 nM   Synthetic analogue   Dd2, 27.5 nM   Dd2, 27.5 nM   Synthetic analogue   Dd2, 27.8 nM   Synthetic analogue   Dd2, 27.8 nM   Synthetic analogue   Dd2, 200.2 nM   Dd2, 200.2 nM   Synthetic analogue   Dd2, 36.7 nM   Synthetic analogue   Synthetic analogu |             |                    | Dd2, 12.9 nM        |                             |                          |
| 27  | 26          | Compound 25b       | 3D7, 4.4 nM         | Synthetic analogue          |                          |
| Dd2, 76.0 nM   Synthetic analogue   |             |                    | Dd2, 6.5 nM         |                             |                          |
| 28  | 27          | Compound 25c       | 3D7, 47.8 nM        | Synthetic analogue          |                          |
| Dd2, 17.8 nM   Synthetic analogue   Dd2, 27.5 nM  |             |                    | Dd2, 76.0 nM        |                             |                          |
| 29         Compound 25e         3D7, 28.1 nM Dd2, 27.5 nM         Synthetic analogue           30         Compound 28a         3D7, 34.4 nM Dd2, 71.8 nM         Synthetic analogue           31         Compound 28b         3D7, 57.6 nM Dd2, 200.2 nM         Synthetic analogue           32         Compound 28c         3D7, 355.7 nM Dd2, 300.3 nM         Synthetic analogue           33         Compound 28d         3D7, 230.3 nM Dd2, 296.7 nM         Synthetic analogue           34         Compound 28e         3D7, 314.2 nM Dd2, 215.7 nM Dd2, 421.5 nM         Synthetic analogue           35         Compound 34a         3D7, 47.9 nM Dd2, 36.7 nM         Synthetic analogue           36         Compound 34b         3D7, 452.0 nM Dd2, 346.5 nM         Synthetic analogue           37         Compound 34c         3D7, 177.5 nM Dd2, 287.8 nM         Synthetic analogue           38         Compound 34d         3D7, 302.4 nM Dd2, 388.4 nM         Synthetic analogue           39         Compound 35a         3D7, 303.5 nM Dd2, 305.1 nM         Synthetic analogue           40         Compound 35b         3D7, 428.5 nM Dd2, 305.1 nM         Synthetic analogue           41         Compound 35e         3D7, 287.4 nM Dd2, 196.7 nM         Synthetic analogue           42         Octaminomycin A         3D7, 1.5 μM  | 28          | Compound 25d       | 3D7, 13.4 nM        | Synthetic analogue          |                          |
| Dd2, 27.5 nM   Synthetic analogue   |             |                    | Dd2, 17.8 nM        |                             |                          |
| 30       Compound 28a       3D7, 34.4 nM       Synthetic analogue         31       Compound 28b       3D7, 57.6 nM       Synthetic analogue         32       Compound 28c       3D7, 355.7 nM       Synthetic analogue         33       Compound 28d       3D7, 355.7 nM       Synthetic analogue         34       Compound 28e       3D7, 314.2 nM       Synthetic analogue         35       Compound 34a       3D7, 47.9 nM       Synthetic analogue         36       Compound 34b       3D7, 452.0 nM       Synthetic analogue         37       Compound 34c       3D7, 177.5 nM       Synthetic analogue         38       Compound 34d       3D7, 362.4 nM       Synthetic analogue         39       Compound 35a       3D7, 303.5 nM       Synthetic analogue         40       Compound 35b       3D7, 428.5 nM       Synthetic analogue         40       Compound 35e       3D7, 287.4 nM       Synthetic analogue         41       Compound 35e       3D7, 287.4 nM       Synthetic analogue         Dd2, 166 μM       Dd2, 196.7 nM       Synthetic analogue  | 29          | Compound 25e       | 3D7, 28.1 nM        | Synthetic analogue          |                          |
| Dd2, 71.8 nM   Synthetic analogue   |             |                    | Dd2, 27.5 nM        |                             |                          |
| 31         Compound 28b         3D7, 57.6 nM Dd2, 200.2 nM         Synthetic analogue           32         Compound 28c         3D7, 355.7 nM Synthetic analogue         Synthetic analogue           33         Compound 28d         3D7, 230.3 nM Dd2, 256.7 nM         Synthetic analogue           34         Compound 28e         3D7, 314.2 nM Dd2, 314.2 nM Dd2, 367. nM         Synthetic analogue           35         Compound 34a         3D7, 47.9 nM Synthetic analogue         Synthetic analogue           36         Compound 34b         3D7, 452.0 nM Dd2, 346.5 nM         Synthetic analogue           37         Compound 34c         3D7, 177.5 nM Dd2, 287.8 nM         Synthetic analogue           38         Compound 34d         3D7, 362.4 nM Synthetic analogue         Synthetic analogue           39         Compound 35a         3D7, 303.5 nM Synthetic analogue         Synthetic analogue           40         Compound 35b         3D7, 428.5 nM Synthetic analogue           40         Compound 35e         3D7, 287.4 nM Synthetic analogue           41         Compound 35e         3D7, 287.4 nM Synthetic analogue           42         Octaminomycin A Dd2, 1.6 μM         Strentomyces sp. RK85-270 Dange et al. (2017)   | 30          | Compound 28a       | 3D7, 34.4 nM        | Synthetic analogue          |                          |
| Dd2, 200.2 nM   Synthetic analogue   Dd2, 300.3 nM   Synthetic analogue   Dd2, 300.3 nM   Synthetic analogue   Dd2, 256.7 nM   Dd2, 256.7 nM   Synthetic analogue   Dd2, 256.7 nM   Dd2, 421.5 nM   Synthetic analogue   Dd2, 421.5 nM   Synthetic analogue   Dd2, 366.7 nM   Synthetic analogue   Dd2, 366.7 nM   Synthetic analogue   Dd2, 346.5 nM   Synthetic analogue   Dd2, 346.5 nM   Synthetic analogue   Dd2, 287.8 nM   Synthetic analogue   Dd2, 318.4 nM   Synthetic analogue   Dd2, 260.8 nM   Synthetic analogue   Dd2, 305.1 nM   Synthetic analogue   Dd2, 305.1 nM   Synthetic analogue   Dd2, 305.1 nM   Synthetic analogue   Dd2, 196.7 nM   Dd2, 1.6 μM   Strentomyces SD, RK85-270   Jang et al. (2017)   |             |                    | Dd2, 71.8 nM        |                             |                          |
| 32         Compound 28c         3D7, 355.7 nM Dd2, 300.3 nM         Synthetic analogue         Kiefer et al. (2019)           33         Compound 28d         3D7, 230.3 nM Dd2, 256.7 nM         Synthetic analogue         Kiefer et al. (2019)           34         Compound 28e         3D7, 314.2 nM Dd2, 421.5 nM         Synthetic analogue           35         Compound 34a         3D7, 47.9 nM Dd2, 36.7 nM         Synthetic analogue           36         Compound 34b         3D7, 452.0 nM Dd2, 346.5 nM         Synthetic analogue           37         Compound 34c         3D7, 177.5 nM Dd2, 287.8 nM         Synthetic analogue           38         Compound 34d         3D7, 362.4 nM Synthetic analogue         Synthetic analogue           Dd2, 318.4 nM         Synthetic analogue           39         Compound 35a         3D7, 303.5 nM Synthetic analogue           Dd2, 260.8 nM         Synthetic analogue           40         Compound 35b         3D7, 428.5 nM Synthetic analogue           Dd2, 305.1 nM         Synthetic analogue           41         Compound 35e         3D7, 287.4 nM Synthetic analogue           Dd2, 166.7 nM         Dd2, 1.6 \( \alpha M \) Streptomyces sp. RK85-270         Jang et al. (2017)  | 31          | Compound 28b       | 3D7, 57.6 nM        | Synthetic analogue          |                          |
| Dd2, 300.3 nM   Synthetic analogue   Synthetic analogue   Dd2, 256.7 nM     34  |             |                    | Dd2, 200.2 nM       |                             |                          |
| 33 Compound 28d 3D7, 230.3 nM Dd2, 256.7 nM  34 Compound 28e 3D7, 314.2 nM Synthetic analogue  35 Compound 34a 3D7, 47.9 nM Synthetic analogue  36 Compound 34b 3D7, 452.0 nM Synthetic analogue  37 Compound 34c 3D7, 177.5 nM Synthetic analogue  38 Compound 34d 3D7, 362.4 nM Synthetic analogue  39 Compound 35a 3D7, 303.5 nM Synthetic analogue  39 Compound 35a 3D7, 303.5 nM Synthetic analogue  302, 260.8 nM  40 Compound 35e 3D7, 428.5 nM Synthetic analogue  3D7, 287.4 nM Synthetic analogue  3D7, 287.4 nM Synthetic analogue  3D8, 305.1 nM  41 Compound 35e 3D7, 287.4 nM Synthetic analogue  3D8, 305.1 nM  42 Octaminomycin A 3D7, 1.5 μM  3D9, 1.6 μM  3D7, 1.5 μM  3D8, 287.4 nM Synthetic analogue  3D9, 287.4 nM Synthetic analogue  3D9, 287.4 nM Synthetic analogue  3D9, 1.5 μM  3D1, 1.5 μM  3D2, 1.6 μM  3D3, 1.6 μM  3D3, 1.6 μM  3D4, 1.6 μM  3D5, 1.5 μM  3D7, 1.5 μM  | 32          | Compound 28c       | 3D7, 355.7 nM       | Synthetic analogue          |                          |
| 33 Compound 28d 3D7, 230.3 nM Dd2, 256.7 nM  34 Compound 28e 3D7, 314.2 nM Synthetic analogue  Dd2, 421.5 nM  35 Compound 34a 3D7, 47.9 nM Synthetic analogue  Dd2, 36.7 nM  36 Compound 34b 3D7, 452.0 nM Synthetic analogue  Dd2, 346.5 nM  37 Compound 34c 3D7, 177.5 nM Synthetic analogue  Dd2, 287.8 nM  38 Compound 34d 3D7, 362.4 nM Synthetic analogue  Dd2, 318.4 nM  39 Compound 35a 3D7, 303.5 nM Synthetic analogue  Dd2, 260.8 nM  40 Compound 35b 3D7, 428.5 nM Synthetic analogue  Dd2, 305.1 nM  41 Compound 35e 3D7, 287.4 nM Synthetic analogue  Dd2, 196.7 nM  42 Octaminomycin A 3D7, 1.5 μM  Dd2, 1.6 μM Streptomyces sp. RK85-270 Jang et al. (2017)   |             |                    | Dd2, 300.3 nM       |                             | Kiefer et al. (2019)     |
| 34       Compound 28e       3D7, 314.2 nM<br>Dd2, 421.5 nM       Synthetic analogue         35       Compound 34a       3D7, 47.9 nM<br>Dd2, 36.7 nM       Synthetic analogue         36       Compound 34b       3D7, 452.0 nM<br>Dd2, 346.5 nM       Synthetic analogue         37       Compound 34c       3D7, 177.5 nM<br>Dd2, 287.8 nM       Synthetic analogue         38       Compound 34d       3D7, 362.4 nM<br>Dd2, 318.4 nM       Synthetic analogue         39       Compound 35a       3D7, 303.5 nM<br>Dd2, 260.8 nM       Synthetic analogue         40       Compound 35b       3D7, 428.5 nM<br>Dd2, 305.1 nM       Synthetic analogue         41       Compound 35e       3D7, 287.4 nM<br>Dd2, 196.7 nM       Synthetic analogue         42       Octaminomycin A       3D7, 1.5 μM<br>Dd2, 1.6 μM       Streptomyces sp. RK85-270       Jane et al. (2017)  | 33          | Compound 28d       |                     | Synthetic analogue          | ()                       |
| Dd2, 421.5 nM       Dd2, 421.5 nM       Synthetic analogue         35 Compound 34a       3D7, 47.9 nM       Synthetic analogue         36 Compound 34b       3D7, 452.0 nM       Synthetic analogue         37 Compound 34c       3D7, 177.5 nM       Synthetic analogue         38 Compound 34d       3D7, 362.4 nM       Synthetic analogue         39 Compound 35a       3D7, 303.5 nM       Synthetic analogue         40 Compound 35b       3D7, 428.5 nM       Synthetic analogue         40 Compound 35e       3D7, 428.5 nM       Synthetic analogue         41 Compound 35e       3D7, 287.4 nM       Synthetic analogue         42 Octaminomycin A       3D7, 1.5 μM         Dd2, 1.6 μM       Streptomyces sp. RK85-270       Jang et al. (2017)   |             |                    |                     |                             |                          |
| 35 Compound 34a 3D7, 47.9 nM Dd2, 36.7 nM 36 Compound 34b 3D7, 452.0 nM Synthetic analogue Dd2, 346.5 nM 37 Compound 34c 3D7, 177.5 nM Synthetic analogue Dd2, 287.8 nM 38 Compound 34d 3D7, 362.4 nM Synthetic analogue Dd2, 318.4 nM 39 Compound 35a 3D7, 303.5 nM Synthetic analogue Dd2, 260.8 nM 40 Compound 35b 3D7, 428.5 nM Synthetic analogue Dd2, 305.1 nM 41 Compound 35e 3D7, 287.4 nM Synthetic analogue Dd2, 196.7 nM 42 Octaminomycin A 3D7, 1.5 μM Dd2, 1.6 μM Streptomyces sp. RK85-270 Jang et al. (2017)   | 34          | Compound 28e       |                     | Synthetic analogue          |                          |
| Dd2, 36.7 nM         36 Compound 34b       3D7, 452.0 nM       Synthetic analogue         37 Compound 34c       3D7, 177.5 nM       Synthetic analogue         38 Compound 34d       3D7, 362.4 nM       Synthetic analogue         Dd2, 318.4 nM       Synthetic analogue         39 Compound 35a       3D7, 303.5 nM       Synthetic analogue         Dd2, 260.8 nM       Synthetic analogue         40 Compound 35b       3D7, 428.5 nM       Synthetic analogue         Dd2, 305.1 nM       Synthetic analogue         41 Compound 35e       3D7, 287.4 nM       Synthetic analogue         Dd2, 196.7 nM       Dd2, 196.7 nM         42 Octaminomycin A       3D7, 1.5 μM         Dd2, 1.6 μM       Streptomyces sp. RK85-270       Jang et al. (2017)   |             |                    |                     |                             |                          |
| 36       Compound 34b       3D7, 452.0 nM       Synthetic analogue         37       Compound 34c       3D7, 177.5 nM       Synthetic analogue         38       Compound 34d       3D7, 362.4 nM       Synthetic analogue         39       Compound 35a       3D7, 303.5 nM       Synthetic analogue         40       Compound 35b       3D7, 428.5 nM       Synthetic analogue         41       Compound 35e       3D7, 287.4 nM       Synthetic analogue         41       Compound 35e       3D7, 287.4 nM       Synthetic analogue         42       Octaminomycin A       3D7, 1.5 μM         Dd2, 1.6 μM       Streptomyces sp. RK85-270       Jang et al. (2017)  | 35          | Compound 34a       |                     | Synthetic analogue          |                          |
| Dd2, 346.5 nM 3D7, 177.5 nM Dd2, 287.8 nM 38 Compound 34d 3D7, 362.4 nM Dd2, 318.4 nM 39 Compound 35a Dd2, 260.8 nM 40 Compound 35b Dd2, 305.1 nM 41 Compound 35e Dd2, 397, 428.5 nM Dd2, 305.1 nM AD2, 16.6 μM  Synthetic analogue Synthetic analogue Dd2, 305.1 nM  Synthetic analogue  |             |                    |                     |                             |                          |
| 37 Compound 34c 3D7, 177.5 nM Dd2, 287.8 nM 38 Compound 34d 3D7, 362.4 nM Synthetic analogue Dd2, 318.4 nM 39 Compound 35a 3D7, 303.5 nM Synthetic analogue Dd2, 260.8 nM 40 Compound 35b 3D7, 428.5 nM Synthetic analogue Dd2, 305.1 nM 41 Compound 35e 3D7, 287.4 nM Synthetic analogue Dd2, 196.7 nM 42 Octaminomycin A 3D7, 1.5 μM Dd2, 1.6 μM Streptomyces sp. RK85-270 Jang et al. (2017)   | 36          | Compound 34b       |                     | Synthetic analogue          |                          |
| Dd2, 287.8 nM  3B Compound 34d  3D7, 362.4 nM  Dd2, 318.4 nM  3P Compound 35a  3D7, 303.5 nM  Dd2, 260.8 nM  40 Compound 35b  3D7, 428.5 nM  Dd2, 305.1 nM  41 Compound 35e  3D7, 287.4 nM  Dd2, 196.7 nM  42 Octaminomycin A  3D7, 1.5 μM  Dd2, 1.6 μM  Synthetic analogue  Synthetic analogue  Synthetic analogue  Synthetic analogue  Dd2, 196.7 nM  Synthetic analogue  Dd2, 196.7 nM  Streptomyces sp. RK85-270  Jang et al. (2017)  | 25          | 6 12/              |                     | C 1 : 1                     |                          |
| 38       Compound 34d       3D7, 362.4 nM       Synthetic analogue         39       Compound 35a       3D7, 303.5 nM       Synthetic analogue         40       Compound 35b       3D7, 428.5 nM       Synthetic analogue         Dd2, 305.1 nM       Synthetic analogue         41       Compound 35e       3D7, 287.4 nM       Synthetic analogue         Dd2, 196.7 nM         42       Octaminomycin A       3D7, 1.5 μM         Dd2, 1.6 μM       Streptomyces sp. RK85-270       Jang et al. (2017)  | 3/          | Compound 34c       |                     | Synthetic analogue          |                          |
| Dd2, 318.4 nM 3D7, 303.5 nM Dd2, 260.8 nM  40 Compound 35b 3D7, 428.5 nM Dd2, 305.1 nM  41 Compound 35e 3D7, 287.4 nM Dd2, 196.7 nM  42 Octaminomycin A 3D7, 1.5 μM Dd2, 1.6 μM  Synthetic analogue Synthetic analogue Dd2, 196.7 nM  | 20          | C 12/1             |                     | C .1 .: 1                   |                          |
| 39 Compound 35a 3D7, 303.5 nM Synthetic analogue Dd2, 260.8 nM  40 Compound 35b 3D7, 428.5 nM Synthetic analogue Dd2, 305.1 nM  41 Compound 35e 3D7, 287.4 nM Synthetic analogue Dd2, 196.7 nM  42 Octaminomycin A 3D7, 1.5 μM Dd2, 1.6 μM Streptomyces sp. RK85-270 Jang et al. (2017)   | 26          | Compound 34d       |                     | Synthetic analogue          |                          |
| Dd2, 260.8 nM  40 Compound 35b 3D7, 428.5 nM Synthetic analogue Dd2, 305.1 nM  41 Compound 35e 3D7, 287.4 nM Synthetic analogue Dd2, 196.7 nM  42 Octaminomycin A 3D7, 1.5 μM Dd2, 1.6 μM Streptomyces sp. RK85-270 Jang et al. (2017)  | 20          | Compound 25a       |                     | Symphopia analogua          |                          |
| 40       Compound 35b       3D7, 428.5 nM       Synthetic analogue         Dd2, 305.1 nM       Dd2, 305.1 nM         41       Compound 35e       3D7, 287.4 nM       Synthetic analogue         Dd2, 196.7 nM       Dd2, 196.7 nM         42       Octaminomycin A       3D7, 1.5 μM         Dd2, 1.6 μM       Streptomyces sp. RK85-270       Jang et al. (2017)   | 39          | Compound 37a       |                     | Synthetic analogue          |                          |
| Dd2, 305.1 nM  41 Compound 35e 3D7, 287.4 nM Synthetic analogue Dd2, 196.7 nM  42 Octaminomycin A 3D7, 1.5 μM Dd2, 1.6 μM Streptomyces sp. RK85-270 Jang et al. (2017)  | 40          | Compound 35h       |                     | Synthetic analogue          |                          |
| <ul> <li>41 Compound 35e 3D7, 287.4 nM Synthetic analogue</li></ul>   | 10          | Compound 370       |                     | Synthetic analogue          |                          |
| Dd2, 196.7 nM  42 Octaminomycin A $3D7$ , 1.5 $\mu$ M  Dd2, 1.6 $\mu$ M  Streptomyces sp. RK85-270 lang et al. (2017)   | 41          | Compound 35e       |                     | Synthetic analogue          |                          |
| 42 Octaminomycin A 3D7, 1.5 μM  Dd2, 1.6 μM  Streptomyces sp. RK85-270 Jang et al. (2017)   |             | Compound 37c       |                     | by minetic analogue         |                          |
| Dd2, 1.6 $\mu$ M Streptomyces sp. RK85-270 Jang et al. (2017)   | 42          | Octaminomycin A    |                     |                             |                          |
| Streptomyces SD. KK8 3-2/U lang et al. (2017)   |             |                    |                     |                             |                          |
| $K1, 1.3 \mu l V l$   |             |                    | K1, 1.3 $\mu$ M     | Streptomyces sp. RK85-270   | Jang et al. (2017)       |
| 43 Octaminomycin B 3D7, 1.5 $\mu$ M   | 43          | Octaminomycin B    |                     |                             |                          |
| Dd2, 1.1 μM   |             |                    |                     |                             |                          |
| K1, $0.85 \mu M$  |             |                    | •                   |                             |                          |



role in the activity. Furthermore, the increasing of sugar units slightly reduced the potency of the tested compounds. In addition to that four C-glycosylated benz[ $\alpha$ ]anthraquinones, four other anthraquinones have been obtained from the culture of the terrestrial *Streptomyces* sp. BCC27095 and identified as steffimycins B (**48**) and C (**49**), 10-dihydrosteffimycin B (**50**) and 7-deoxysteffimycinone (**51**). However, compound **49** showed strong activity against *P. falciparum* K1 with IC<sub>50</sub> of 0.53  $\mu$ M, while compounds **48**, **50** and **51** demonstrated a good activity with IC<sub>50</sub> values of 4.76  $\mu$ M, 2.19  $\mu$ M and 8.03  $\mu$ M, respectively (Intaraudom et al., 2015).

#### 3.4. Geldanamycin analogues

Another important class of antiplasmodial compounds from *Streptomyces* is composed of geldanamycin (52) and its congeners 17-odemethylgeldanamycin (53), 17demethoxyreblastatin (54) isolated from the terrestrial Streptomyces sp. BCC71188, as well as herbimycin G (55) obtained from the marine Streptomyces sp. USC-16018 (Supong et al. 2016b; ) (Buedenbender et al., 2018). Geldanamycin (52) and 17-demethoxyreblastatin (54) gave similar strong potencies of IC<sub>50</sub> values 0.35  $\mu$ g/ml and 0.31  $\mu$ g/ml, respectively, against P. falciparum K1, while 17-odemethylgeldanamycin (54) was moderately active with an IC<sub>50</sub> value of 1.90  $\mu$ g/ml against the same parasitic strain (Table 5). More recently, the research works carried out by Buedenbender et al. (2018) dealt with the identification of herbimycin G (55) containing a geldanamycin scaffold in its structure. Compound 55 showed at 40  $\mu$ M, 77.2% inhibition of P. falciparum 3D7 and 81.7% inhibition of P. falciparum Dd2 (Buedenbender et al., 2018). The structures of the antiplasmodial geldanamycin derivatives are presented in Figure 5.

# 3.5. Pactamycin analogues

Pactamycin (56) is a well-known antibiotic discovered by the Upjohn Company in the early 1960s and has been reported during the last decades from Streptomyces pactum ATCC 27456 by Almabruk et al. (2013). During their investigations, the authors also isolated two other pactamycin analogues indexed TM-025 (57) and TM-026 (58) (Figure 6) which were submitted to mutasynthetic strategy to generate their fluorinated derivatives TM-025F (59) and TM-026F (60), respectively. All five compounds were screened for their antiplasmodial activity against three P. falciparum strains including the chloroquinesensitive strain D6 and two multidrug-resistant strains Dd2 and 7G8. The results (Table 6) showed that pactamycin (56) was the most active compound against the three strains, while its congeners compounds 57 and 58, as well as their fluorinated derivatives 59 and 60, were slightly less active against the three strains with IC<sub>50</sub> values ranging from 3.9 nM to 39.1 nM which remains in an excellent range of activity for all the compounds (Almabruk et al., 2013).



From the literature accessed during our survey, three compounds described as polyethers have been isolated from *Streptomyces* sp. and described as demonstrating an antiplasmodial activity (Table 7, Figure 7). Abierixin (**61**) and its methylated derivative 29-O-methylabierixin (**62**) were isolated from *Streptomyces* sp. BCC72023 showed a moderate potency against *P. falciparum* K1 with IC<sub>50</sub> values of 2.58  $\mu$ g/ml and 1.40  $\mu$ g/ml, respectively, while compound **63** isolated from *Streptomyces* sp. H668 was active against *P. falciparum* D6 and W2 with IC<sub>50</sub> ranging from 100 to 200 ng/ml (Na et al., 2008; Supong, Thawai, et al., 2016).

#### 3.7. Diketopiperazines

Two diketopiperazines (**64** and **65**) and one dimeric diketopiperazine (**66**) have been reported as antiplasmodial agents from *Streptomyces* strains (Table 8, Figure 8). Thus, the chemical investigations of the isolate S1 of *Streptomyces* sp. led to the isolation of piperafizine A (**64**) demonstrating activity of IC<sub>50</sub> equal to 6.57  $\mu$ M against *P. falciparum* Dd2 (Rakotondraibe et al., 2015), whereas another diketopiperazine identified as L-Pro-L-Leu (**65**) was obtained from *Streptomyces* sp. USC-16018 with 45.9 % inhibition of *P. falciparum* 3D7 and 39.0 % inhibition of *P. falciparum* Dd2 at 40  $\mu$ M (Buedenbender et al., 2018). A dimeric analogue called naseseazine C (**66**) was reported in 2016 from the culture of *Streptomyces* sp. and was moderately active against *P. falciparum* 3D7 with an IC<sub>50</sub> of 3.52  $\mu$ M (Buedenbender et al., 2016).

# 3.8. Miscelleaneous

Several other specialised metabolites from Streptomyces have been identified as antiplasmodial principles (Table 9, Figure 9). Among them, farneside A (67) was reported as a sesquiterpenoid nucleoside ether from *Streptomyces* sp. CNT-372 with moderate activity (IC<sub>50</sub> of 69.3  $\mu$ M) against *P. falciparum* 3D7 while 2methylthio- $N^7$ -methyl-cis-zeatin (68), the first  $N^7$ -methylated zeatin-type natural product has been reported from *Streptomyces* 80H647 with a GI<sub>50</sub> of 2.4  $\mu$ M against the same parasite (Ilan et al., 2013; Lopez et al., 2000). One polyenoic acid amide natural product named annimycin B (69) and a metabolite containing a  $\gamma$ -butyrolactone and 2-hydroxy-3formylaminobenzoic acid moieties called opantimycin A (70) showed weak activity with 30% inhibition of P. falciparum Dd2, HB3, 3D7 at 2.5  $\mu$ M; and IC<sub>50</sub> of 13  $\mu$ g/ml against P. falciparum 3D7, respectively (Zhang et al. 2014; ) (Nogawa et al., 2017).

One benzoisochromanequinone frenolicin B (71) obtained from *Streptomyces roseofulvus* displayed good antiplasmodial activity against *P. falciparum* HB3, Dd2 and 7G8 with IC<sub>50</sub> values of 600 nM, 800 nM and 800 nM, respectively (Fitzgerald et al., 2011). Similarly, the carbazole antibiotics named carbazomycins B (72) and C (73) were obtained from *Streptomyces* sp. BCC27095, as well as the tripyrrole pigment metacycloprodigiosin (74) and the nitrophenyl-substituted polyketide spectinabilin (75) from *Streptomyces spectabilis* 



Figure 4. Antiplasmodial anthraquinones from Streptomyces

 Table 4

 Antiplasmodial anthraquinones from Streptomyces.

| N°         | Name                    | Strain, IC 50         | Source                    | Reference                |
|------------|-------------------------|-----------------------|---------------------------|--------------------------|
| 44         | Urdamycinone E          | K1, 0.0534 $\mu$ g/ml |                           |                          |
| 45         | Urdamycinone G          | K1, $0.142 \mu g/ml$  | Streptomyces sp. BCC45596 | Supong et al. (2012)     |
| 46         | dehydroxyaquayamycin    | K1, 2.93 $\mu$ g/ml   | Streptomytes sp. BCC4))90 | Supong et al. (2012)     |
| <b>4</b> 7 | urdamycin E             | K1, 0.173 $\mu$ g/ml  |                           |                          |
| 48         | steffimycin B           | K1, 2.19 $\mu$ M      |                           |                          |
| 49         | Steffimycin C           | K1, 0.53 $\mu$ M      | Company of BCC27005       | Intaraudom et al. (2015) |
| 50         | 10-Dihydrosteffimycin B | K1, $4.76 \mu M$      | Streptomyces sp. BCC27095 |                          |
| 51         | 7-deoxysteffimycinone   | K1, 8.03 $\mu$ M      |                           |                          |

Figure 5. Antiplasmodial geldanamycin analogues from Streptomyces.

 Table 5

 Antiplasmodial geldanamycin analogues from Streptomyces.

| <b>N</b> ° | Name                     | Strain, IC 50                             | Source                     |                                      |
|------------|--------------------------|---|----------------------------|--------------------------------------|
| 52         | Geldanamycin             | K1, 0.35 $\mu$ g/ml                       |                            | 6 6 1 1 1                            |
| 53         | 17-Odemethylgeldanamycin | K1, 1.90 $\mu$ g/ml                       | Streptomyces sp. BCC71188  | Supong, Sripreechasak, et al. (2016) |
| 54         | 17-demethoxyreblastatin  | K1, 0.31 $\mu$ g/ml                       |                            | (2010)                               |
| 55         | Herbimycin G             | 3D7, 77.2 % Inhib. at 40 $\mu\mathrm{M}$  | Streptomyces sp. USC-16018 | Buedenbender et al. (2018)           |
|            |                          | Dd2, 81.7 % Inhib. at 40 $\mu \mathrm{M}$ |                            |                                      |



Figure 6. Antiplasmodial pactamycin analogues from *Streptomyces*.

 Table 6

 Antiplasmodial pactamycin analogues from Streptomyces.

| N° | Name       | Strain, IC 50 | Source                         | Reference                 |
|----|------------|---------------|--------------------------------|---------------------------|
| 56 | Pactamycin | D6, 4.9 nM    | Streptomyces pactum ATCC 27456 |                           |
|    |            | Dd2, 3.9 nM   |                                |                           |
|    |            | 7G8, 4.2 nM   |                                |                           |
| 57 | TM-025     | D6, 7.7 nM    | Streptomyces pactum ATCC 27456 |                           |
|    |            | Dd2, 10.5 nM  |                                |                           |
|    |            | 7G8, 7.1 nM   |                                | A1 1 1                    |
| 58 | TM-026     | D6, 11.5 nM   | Streptomyces pactum ATCC 27456 | Almabruk<br>et al. (2013) |
|    |            | Dd2, 14.0 nM  |                                | ct al. (2013)             |
|    |            | 7G8, 9.1 nM   |                                |                           |
| 59 | TM-025F    | D6, 12.3 nM   | Synthetic                      |                           |
|    |            | Dd2, 16.8 nM  |                                |                           |
|    |            | 7G8, 12.4 nM  |                                |                           |
| 60 | TM-026F    | D6, 26.5 nM   | Synthetic                      |                           |
|    |            | Dd2, 39.1 nM  |                                |                           |
|    |            | 7G8, 24.9 nM  |                                |                           |

Figure 7. Antiplasmodial polyethers from Streptomyces.

**Table 7** Antiplasmodial polyethers from *Streptomyces*.

| N° | Name                 | Strain, IC 50              | Source                | Reference        |
|----|----------------------|----------------------------|-----------------------|------------------|
| 61 | abierixin            | K1, 2.58 $\mu$ g/ml        | Streptomyces sp.      | Supong, Thawai,  |
| 62 | 29-O-methylabierixin | K1, 1.40 $\mu$ g/ml        | BCC72023              | et al. (2016)    |
| 63 | //                   | D6 and W2, 100 - 200 ng/ml | Streptomyces sp. H668 | Na et al. (2008) |



Figure 8. Antiplasmodial diketopiperazines from *Streptomyces*.

**Table 8** 

 Antiplasmodial diketopiperazines from *Streptomyces*.

| N° | Name           | Strain, IC 50   | Source                     | Reference                   |
|----|----------------|---|----------------------------|-----------------------------|
| 64 | Piperafizine A | Dd2, 6.57 $\mu$ M   | Streptomyces isolate S.1   | Rakotondraibe et al. (2015) |
| 65 | L-Pro-L-Leu    | 3D7, 45.9 % Inhib. at 40 $\mu$ M Dd2, 39.0 % Inhib. at 40 | Streptomyces sp. USC-16018 | Buedenbender et al. (2018)  |
| 6  | Naseseazine C  | μΜ<br>3D7, 3.52 $μ$ Μ                                     | Streptomyces sp.           | Buedenbender et al. (2016)  |

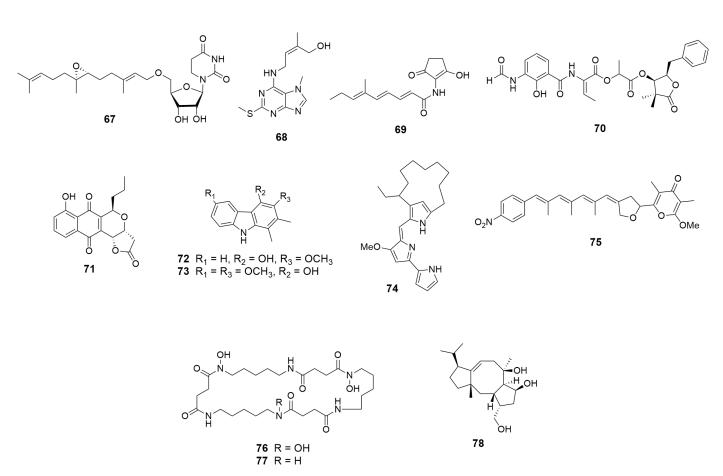


Figure 9. Other antiplasmodial compounds from Streptomyces.



 Table 9

 Other antiplasmodial compounds from Streptomyces.

| N°        | Name                                   | Strain, IC 50                                  | Source                     | Reference                               |  |
|-----------|--|--|----------------------------|---|--|
| 67        | Farneside A                            | 3D7, 69.3 $\mu$ M                              | Streptomyces sp. CNT-372   | Ilan et al. (2013)                      |  |
| 68        | 2-Methylthio- $N^7$ -methyl-cis-zeatin | 3D7, GI50 2.4 $\mu$ M                          | Streptomyces sp. 80H647    | Lopez et al. (2000)                     |  |
| 69        | Annimycin B                            | Dd2, HB3 and 3D7, 30% of inhib. at 2.5 $\mu M$ | S. asterosporus DSM 41452  | Zhang et al. (2018)                     |  |
| <b>70</b> | Opantimycin A                          | 3D7, 13 μg/ml                                  | Streptomyces sp. RK88-1355 | Nogawa et al. (2017)                    |  |
| 71        | Frenolicin B                           | HB3, 600 nM                                    | Streptomyces roseofulvus   | Fitzgerald et al. (2011)                |  |
|           |  | Dd2, 800 nM                                    |                            |   |  |
|           |  | 7G8, 800 nM                                    |                            |   |  |
| 72        | Carbazomycin B                         | K1, 2.37 $\mu$ g/ml                            | Streptomyces sp. BCC26924  | Intaraudom et al. (2011)                |  |
| 73        | Carbazomycin C                         | K1, 2.10 μg/ml                                 | Streptomyces sp. BCC20924  |   |  |
| <b>74</b> | Metacycloprodigiosin                   | K1, $0.0050 \mu g/ml$                          | C                          | Il1 (2002)                              |  |
| 75        | Spectinabilin                          | K1, 7.8 $\mu$ g/ml                             | S. spectabilis BCC 4785    | Isaka et al. (2002)                     |  |
| 76        | Nocardamine                            | K1, 3.20 μg/ml                                 |                            | 6 6 1 1                                 |  |
| 77        | Dehydroxynocardamine                   | K1, 2.63 µg/ml                                 | Streptomyces sp.BCC71188   | Supong, Sripreechasak,<br>et al. (2016) |  |
| <b>78</b> | Cyclooctatin                           | K1, $7.14 \mu g/ml$                            |                            | et al. (2010)                           |  |

BCC4785, were all evaluated against *P. falciparum* K1 and only compound **74** gave a strong activity with IC<sub>50</sub> of 0.0050  $\mu$ g/ml while compounds **72** and **73** gave a moderate activity with IC<sub>50</sub> values of 2.37  $\mu$ g/ml and 2.10  $\mu$ g/ml, respectively; and finally, compound **75** was the less active with an IC<sub>50</sub> of 7.8  $\mu$ g/ml (Intaraudom et al., 2011; Isaka et al., 2002).

Two cyclopeptides namely nocardamine (**76**) and its derivative dehydroxynocardamine (**77**) have been isolated and characterized from *Streptomyces* sp. BCC71188. Both compounds were tested against P. falciparum K1 and found to be moderately active with IC $_{50}$  values of 3.20  $\mu$ g/ml and 2.63  $\mu$ g/ml, respectively (Supong, Sripreechasak, et al., 2016). Finally, the diterpenoid cyclooctatin (**78**) was obtained from the chemical investigations of *Streptomyces* sp. BCC71188 gave a moderate activity with an IC $_{50}$  of 7.14  $\mu$ g/ml against *P. falciparum* K1 (Supong, Sripreechasak, et al., 2016).

# 4. CONCLUSION AND FUTURE PROSPECTS

The data provided by the literature on the antiplasmodial compounds from Streptomyces support that these bacteria represent important sources of bioactive metabolites that can be considered as interesting candidates for new drugs discovery. In addition to be well reported as antibiotics, many Streptomycesderived compounds demonstrated strong activities and were in some cases more effective than the reference drugs. For instance, three compounds including the macrolide munumbucin D and the two octadepsipeptides octaminomycins A (42) and B (43) displayed good antiplasmodial activity and did not show any cytotoxicity against several cell lines that increases their pharmacological interest for new drug development. Additional close checking indicated that several other compounds from Streptomyces have displayed strong or good potency against the drug-resistant strain K1 of P. falciparum. Among them, bafilomycin A1 (9), concanamycin A (10), elaiophylin (17), cyclomarin C (23), urdamycinone E (44), geldanamycin (52) and metacycloprodigiosin (74) demonstrated strong antiplasmodial with IC<sub>50</sub> values of 0.041  $\mu$ g/ml, 0.2 nM, 0.22  $\mu$ g/ml, 0.24  $\mu$ g/ml, 0.0534  $\mu$ g/ml, 0.35  $\mu$ g/ml and 0.0050  $\mu$ g/ml, respectively, against P falciparum K1. Inded, the discovery of a new compound with high potency against a chloroquine-resistant strain of P falciparum like K1 or Dd2 might be a good starting point to address the problem of resistance of P falciparum to prescribed drugs which is one of the most important factors to control in the eradication of malaria. Overall, despite the interesting  $in\ vitro$  antiplasmodial activity recorded so far for the indicated compounds, further  $in\ vivo$ , pharmacokinetic or ADMET studies are necessary to obtain more insights on their action mechanism, solubility or toxicity which are important to manufacturing a drug.

# **CONFLICTS OF INTEREST**

The authors have no relevant financial or non-financial interests to disclose.

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